

CRYOSTOR™ QUICK REFERENCE CRYOPRESERVATION PROTOCOL

- 1) Place cells to be cryopreserved into suspension (*mechanical or enzymatic dissociation*)
- 2) Centrifuge cells to obtain cell pellet
- 3) Remove supernatant
Note: Remove as much culture media as possible, to reduce dilution of CryoStor™ solution.
- 4) **ISOLATION:** Add cold (2-8°C) CryoStor™
 - a. Cell concentrations: $0.5-10 \times 10^6$ cells/ml (higher [cell] possible with testing).
 - b. DMSO is pre-mixed in CryoStor™ - no additives are necessary.
- 5) **PRE-FREEZE:** Incubate cell suspension at 2-8°C for approximately 10 minutes
- 6) **NUCLEATION:** Freeze samples at -80°C
 - a. Use a controlled rate freeze (-1°C/min) or similar protocol for most mammalian cell systems.
 - b. The freezing device or isopropanol container should be pre-cooled to 2-8°C.
 - c. Ice nucleation within the sample (seeding) should be initiated at approximately -5°C using either a liquid nitrogen burst program setting on a controlled rate freezer or mechanical agitation (flick or tap) of the cryovial/sample container after approximately 15-20 min. at -80°C.
 - d. Freeze time (-80°C) using isopropanol containers is recommended to be 3-4 hours.
- 7) **STORAGE:** Place samples into storage
 - a. Store samples at liquid nitrogen temperatures (below -130°C).
 - b. Sample storage at -80°C is only recommended for short-term storage (weeks to months).
- 8) **THAWING:** Thaw samples quickly in a 37°C water bath
 - a. Sample thawing should be conducted with gentle swirling of sample until all visible ice has melted. Approximate thaw time for a 1 ml sample in a cryovial is approximately 3 minutes.
 - b. DO NOT allow sample to warm above chilled temperatures (0-10°C). Cryovials should be cool to the touch when removed from bath. Passive thaw is not recommended.
- 9) Dilute cell/CryoStor™ mixture immediately with culture media
 - a. Dilution procedure can be preformed in a single step.
 - b. The dilution media should be between 20°C and 37°C.
 - c. A dilution ratio of 1:10 (sample to media) or greater is recommended.
- 10) Plate cells in appropriate configuration
- 11) Place cells into culture conditions or utilize immediately
- 12) Viability assessment 24-hours post-thaw*
Note: To obtain an accurate measure of cell viability following cryopreservation, assessment should be performed 24 hours post-thaw and compared to non-frozen controls.

*Sample assessment immediately post-thaw with membrane integrity indicators, such as Trypan Blue, for comparative analysis of sample cell yield and viability often results in significant overestimates of cell survival.

Live/Dead fluorescent assays or metabolic assays (MTT or AlamarBlue) are recommended for more accurate viability assessment. Visual inspection of adherent cells and cells "floating" in the media is also recommended.

CRYOPRESERVATION SOLUTIONS

Designed to prepare and preserve cells in ultra low temperature environments (-80°C to -196°C); CryoStor™ provides a safe, protective environment for cells and tissues during the freezing, storage, and thawing process. Through modulating the cellular biochemical response to the cryopreservation process, CryoStor™ provides for enhanced cell viability and functionality while eliminating the need to include serum, proteins or high levels of cytotoxic agents.



BioLife Solutions Technical Support
425-402-1400 | 866-4BIOLIFE (866-246-5433)
support@BioLifeSolutions.com

www.BioLifeSolutions.com



Source Material **BioPreservation** Cell Products